



## REVIEW ARTICLE | OPEN ACCESS

# Ameliorative potential of conditioning on ischemia-reperfusion injury in diabetes

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Diabetes is a serious metabolic disease characterized by hyperglycemia. Diabetes also leads to several long-term secondary complications. Cardiovascular disease is an important complication of diabetes and is a major contributor to morbidity and mortality in diabetic subjects. The discovery of conditioning-induced ischemic or anoxic tolerance has led to the demonstration of the protective potential of conditioning as a treatment strategy to mitigate ischemia-reperfusion injury. Diabetes modulates multiple metabolic pathways and signal transduction cascades. Some of these pathways may overlap with mechanisms that mediate the beneficial effects of conditioning from the body's reaction to a sublethal insult, indicating the possibility of a potential interaction between diabetes and conditioning. Studies demonstrate that diabetes abrogates the ameliorative effect of various forms of conditioning, such as ischemic preconditioning, ischemic postconditioning, remote ischemic conditioning and pharmacological conditioning, on ischemia-reperfusion injury in various animal models. Moreover, drugs used to treat diabetes may have a potential impact on protection afforded by conditioning from ischemic injury. Potential impact of various anti-diabetic drugs on conditioning-induced protection is also discussed. Overall, the literature suggests that a better understanding of the overlap among pathways activated by diabetes and those involved in induction of ischemia tolerance may help identify ideal conditioning paradigms to protect diabetic subjects from ischemic injury.

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The American Diabetes Association defines diabetes mellitus as a “a group of metabolic diseases characterized by hyperglycemia resulting from defects in insulin secretion, insulin action, or both” (American Diabetes Association, 2014). Approximately 30.3 million Americans are suffering from diabetes, and the economic burden due to diabetes was \$320 billion in 2015 (Centers for Disease Control and Prevention, 2017). Chronic diabetes produces long-term detrimental changes in several organs leading to development of cardiovascular diseases, neuropathy, retinopathy and nephropathy, among other secondary complications (Bhalla et al., 2013; American Diabetes Association, 2014; de Ferranti et al., 2014; Kolber and Scrimshaw, 2014; Fox CS, 2015; National Eye Institute, 2015; Afkarian et al., 2016; Huo X, 2016; Tracey et al., 2016; Pop-Busui et al., 2017; American Optometric Association, 2018). Diabetes also increases the prevalence of and mortality resulting from brain ischemia and myocardial injury (Centers for Disease Control and Prevention, 2003; Almdal et al., 2004; Ottenbacher et al., 2004; Kissela et al., 2005; Alegria et al., 2007; Marso et al., 2007).

### Diabetes and ischemia-reperfusion injury

Diabetes is an established risk factor for ischemic stroke and coronary heart disease (Schramm et al., 2008; Spencer et al., 2008). A meta-analysis of 102 prospective studies demonstrated that diabetes causes about a two-fold increase in the risk of ischemic stroke, coronary heart disease, and deaths ascribed to other vascular complications (Emerging Risk Factors Collaboration et al., 2010). Both type 1 diabetes (T1D) and type 2 diabetes (T2D) enhance cardiovascular mortality and morbidity (Fuller et al., 2001; Janghorbani et al., 2007). The pathophysiology of increased progression and risk of cardiovascular diseases in diabetes patients is not well understood. Pronounced increase in risk factors such as dyslipidemia, hypercoagulability, endothelial dysfunction, and inflammation in diabetic patients may increase the risk of cardiovascular diseases (Plutzky and Brown, 2012). This role has been established by several groups (see reviews (Fonseca et al., 2004; Martin-Timon et al., 2014; Leon and Maddox, 2015)). In comparison to non-diabetic subjects, diabetic individuals display enhanced lipid-rich atheroma, macrophage infiltration and formation of thrombus in the coronary arteries (Moreno et al., 2000). Atherosclerotic plaques in diabetic patients have higher levels of macrophages, T lymphocytes, and inflammatory cells (human leukocyte antigen-DR positive cells) (Cipollone et al., 2003). Accelerated atherosclerosis is seen in diabetic individuals (Otsuka et al., 2012). Overall, the above factors lead to increased risk of thrombosis and subsequent tissue ischemia in diabetics. Therefore, there is a need to develop novel therapeutic approaches to mitigate ischemic injury in diabetic subjects. Research efforts worldwide have focused on evaluating numerous potential strategies to salvage tissue from ischemic damage. The discovery of ischemic/pharmacologic preconditioning, postconditioning and remote conditioning has uncovered novel ways to activate multiple cytoprotective signaling pathways to confer protection against ischemic injury (Dave et al., 2001; Dave et al., 2006; Raval et al., 2006a; Raval et al., 2006b; Koronowski et al., 2015; Hausenloy et al., 2016). The goal of this review article is to describe the published literature that has evaluated the efficacy of conditioning in lowering ischemia-reperfusion injury in the diabetic condition.

### Conditioning

Conditioning denotes the “adaptive process of endogenous protection in which small doses of sub-lethal ischemia protects the organism against a lethal ischemic event” (Adstamongkonkul, 2017). The term *conditioning* is broadly used to describe the induction of ischemia tolerance by a group

of various paradigms. These paradigms include ischemic preconditioning, perconditioning, postconditioning, remote conditioning and pharmacological conditioning. The term ischemic preconditioning describes the protective effect induced by episode(s) of brief ischemia, too brief in themselves to cause tissue damage, before an episode of severe ischemia (Przyklenk, 2013). Ischemic preconditioning is a promising approach to lower the extent of ischemic injury in the heart (Murry et al., 1986), brain (Kitagawa et al., 1990), kidney (Lee and Emala, 2000), small intestine (Pajdo et al., 2001), skeletal muscle (Mounsey et al., 1992) and liver (Lloris-Carsi et al., 1993). Various types of stress conditions such as cortical spreading depression (Kobayashi et al., 1995), electroacupuncture (Xiong et al., 2003), mild epileptic insult (Plamondon et al., 1999), thrombin (Masada et al., 2000), hyperbaric oxygen preconditioning (Wada et al., 1996), and exercise (Lennon et al., 2004; Quindry et al., 2005; McGinnis et al., 2015) also induce ischemia tolerance in various organs. Ischemic perconditioning refers to a therapeutic strategy designed to reduce infarct size by application of conditioning stimulus during the ischemic event (Vinten-Johansen and Shi, 2011). Ischemic postconditioning induces tolerance by restoring blood flow to the ischemic tissue in an intermittent manner, before complete reperfusion occurs (after the severe ischemic insult). This type of conditioning is shown to protect heart (Zhao et al., 2003), brain (Zhao et al., 2006), liver (Sun et al., 2004), kidney (Jonker et al., 2016), and intestines (Sengul et al., 2013; Jia et al., 2017) from ischemia-reperfusion injury. In the case of both ischemic preconditioning and postconditioning, the site exposed to the protective stimulus (brief episodes of ischemia or modified reperfusion) as well as severe ischemia are typically the same. However, in the case of remote ischemic conditioning, brief episode(s) of ischemia on a tissue or organ exert a protective effect on a distant tissue or organ subjected to ischemia (Przyklenk et al., 1993; Dave et al., 2006). Remote conditioning includes remote preconditioning, perconditioning and postconditioning when the remote ischemic event is elicited before, during and after the major ischemic event, respectively. Mechanistic understanding of various forms of conditioning have implicated the possibility of artificially (without inducing ischemia) activating mechanisms mediating ischemic conditioning. This type of conditioning is referred to as pharmacological conditioning (Julier et al., 2003; Luca et al., 2011). Below we provide a summary of literature that has used various conditioning paradigms to protect tissues and organs against ischemic injury in animal models of diabetes (Table 1).

### Ischemic preconditioning and protection in diabetes

Ischemic preconditioning is a paradigm that induces protection against ischemic damage in the same tissue or organ (Murry et al., 1986; Kitagawa et al., 1990; Mounsey et al., 1992; Lloris-Carsi et al., 1993; Stagliano et al., 1999; Lee and Emala, 2000; Pajdo et al., 2001; Dave et al., 2005). As mentioned above, an increase in the risk factors for cardiovascular disease in diabetes enhances the risk of ischemia and the extent of ischemic damage. Therefore, to understand the potential therapeutic benefit of preconditioning during diabetes, studies have evaluated the effect of preconditioning in animal models of diabetes. Ischemic preconditioning of kidney (four cycles of 4-min ischemia followed by 11-min reperfusion) did not produce any ameliorative effect on ischemia-reperfusion-induced injury in streptozotocin diabetic rats (diabetes was induced 1 month prior to ischemia) when histological and biochemical markers were used as surrogates for tissue damage (Ozbilgin et al., 2016). A group using a canine model demonstrated that diabetic and acutely hyperglycemic animals display a reduced protective effect of ischemic preconditioning on coronary artery occlusion/reperfusion-induced myocardial infarction (Kersten et al., 1998; Kersten et al., 2000). Ischemic

Table 1: Summary of studies on the effect of diabetes on conditioning.

Type of Conditioning	Diabetic/ Non-diabetic Population	Model of Ischemia	Conditioning Stimulus	Induction of Protection	Reference(s)
Preconditioning	T1D rats	Kidney ischemia <i>in vivo</i>	4 x 4 min ischemia	No	Ozbilgin et al., 2016
	T1D/Hyperglycemic dogs	Myocardial ischemia <i>in vivo</i>	4 x 5 min ischemia	No	Kersten et al., 1998, 2000
	T1D sheep	Myocardial ischemia <i>in vivo</i>	6 x 5 min ischemia	No	del Valle et al., 2003
	Zucker obese rats	Myocardial ischemia <i>in vivo</i>	6 x 5 min ischemia	No	Katakam et al., 2007
	T1D rats	Isolated heart preparation	4 x 5 min ischemia	No	Tosaki et al., 1996
	T1D/Hyperglycemic rabbits	Myocardial ischemia <i>in vivo</i>	1 x 5 min ischemia	No	Ebel et al., 2003
	T1D rats	Isolated heart preparation	1 x 5 min ischemia	No	Bouchard and Lamontagne, 1998
	T2D rats	Isolated heart preparation	4 x 1 min ischemia	No	Kristiansen et al., 2004
	T2D rats	Isolated heart preparation	3 x 5 min ischemia	No	Tsang et al., 2005
	T2D rats	Isolated heart preparation	1 or 3 x 5 min ischemia	No	Hausenloy et al., 2013
	T1D rats	Focal cerebral ischemia-induced brain injury	3 x 10 min ischemia	Yes	Altintas et al., 2016a; 2016b
	Non-diabetic	Organotypic cultures of the hippocampus	Hypoxia/hypoglycemia	Yes	Badaut et al., 2005
	T1D rats	Retinal ischemia	5-min ischemia, weekly repetitions	Yes	Fernandez et al., 2011, 2012
	T1D rats	Intestine and liver ischemia	1 x 10 min ischemia	Yes	Thomaz Neto et al., 2013
Postconditioning	T1D and T2D mice	Isolated heart preparation	3 or 6 x 10 sec ischemia	No	Przyklenk et al., 2011
	T1D rats	Myocardial ischemia <i>in vivo</i>	3 x 20 sec ischemia	No	Drenger et al., 2011
	T2D mice	Myocardial ischemia <i>in vivo</i>	6 x 10 sec ischemia	No	Bouhidel et al., 2008; Zhu et al., 2012
Remote Preconditioning	T1D rats	Myocardial ischemia <i>in vivo</i>	4 x 5 min liver ischemia	No	Hu et al., 2017
	Hyperglycemia	Myocardial ischemia <i>in vivo</i>	3 x 5 min femoral artery and vein occlusion	No	Baranyai et al., 2015
	T1D	Myocardial ischemia <i>in vivo</i>	1 x 15 min femoral artery occlusion	No	Kiss et al., 2014
Diabetic human subjects	Myocardial infarction	Intermittent upper arm ischemia	No	Moretti et al., 2018	
Pharmacological Preconditioning	Hyperglycemia in rabbits	Myocardial ischemia <i>in vivo</i>	Isoflurane postconditioning	No	Raphael et al., 2010
	Hyperglycemia in dogs	Myocardial ischemia <i>in vivo</i>	Isoflurane preconditioning	No	Kehl et al., 2002
	T1D dogs	Myocardial ischemia <i>in vivo</i>	Isoflurane preconditioning	No	Tanaka et al., 2002
	T1D rats	Myocardial ischemia <i>in vivo</i>	Sevoflurane postconditioning	No	Drenger et al., 2011; Tai et al., 2012
	T1D rats	Myocardial ischemia <i>in vivo</i>	Erythropoietin postconditioning	No	Ghaboura et al., 2011
	T1D rats	Myocardial ischemia <i>in vivo</i>	Morphine preconditioning	No	Gross et al., 2007
	T2D rats	Myocardial ischemia <i>in vivo</i>	Erythropoietin and [D-Ala2, D-Leu5]-enkephalin acetate - preconditioning	No	Hotta et al., 2010
	T2D rats	Myocardial ischemia <i>in vivo</i>	Isoflurane-induced preconditioning	No	Matsumoto et al., 2009

T1D: Type 1 diabetes; T2D: Type 2 diabetes.

preconditioning produced by six cycles of 5-min ischemia and a 5-min reperfusion period did not exert any ameliorative effect on stunning in ischemic heart in diabetic sheep when evaluated in both the first and second window of preconditioning. Instead, the first window of preconditioning worsened ischemia-reperfusion-induced myocardial damage as measured in terms of mechanical function (del Valle et al., 2003). While ischemic preconditioning elicited by a cycle of 5 min of ischemia and 5 min of reperfusion decreases infarct size in the myocardium exposed to 30 mins of left coronary artery ligation and 4 hours of reperfusion in heart of Zucker lean rats, this protection is abolished in insulin-resistant Zucker obese rats (Katakam et al., 2007). In addition, long-term diabetes (4 and 8 weeks of streptozotocin-induced diabetes) is also associated with the lack of protective effect of ischemic preconditioning on ventricular fibrillation, tachycardia, cardiac function, and ion shift abnormalities induced by 30-min ischemia / 30-min reperfusion (Tosaki et al., 1996). In an *in vivo* rabbit model of myocardial ischemia, acute hyperglycemia and diabetes attenuated the cardioprotective effect of the second window of ischemic preconditioning (5 min of left descending coronary artery occlusion) against myocardial ischemia (30 min of ischemia and 2 h of reperfusion) when damage was evaluated in terms of the myocardial infarct size (Ebel et al., 2003). Moreover, diabetic inhibition of myocardial preconditioning was refractory to acute insulin treatment (Ebel et al., 2003). This study suggests that simple correction of hyperglycemia is not able to restore the protective effects of ischemic preconditioning.

Ischemic preconditioning with a single episode of ischemic insult involving 5-min ischemia and 10-min reperfusion prevents ischemia-reperfusion-induced endothelial dysfunction in non-diabetic hearts but not in diabetic hearts (Bouchard and Lamontagne, 1998). However, three such episodes of preconditioning produced a similar beneficial effect on the diabetic heart (Bouchard and Lamontagne, 1998). This study suggests that a more extensive preconditioning stimulus is required to condition diabetic hearts. Ischemic preconditioning induced by 4 cycles of 2 min of ischemia and 3 min of reperfusion does not exert any significant effect on the extent of myocardial infarction induced by 50 min of ischemia and 120 min of reperfusion in rat hearts isolated from Zucker diabetic fatty (obese model of T2D) and lean Goto-Kakizaki rats (lean model of T2D) (Kristiansen et al., 2004). Experiments on isolated rat heart preparation observed that while a single short (5-min) episode of ischemic preconditioning elicits a protective effect only on non-diabetic heart, more severe ischemic preconditioning stimuli involving three episodes of 5-min ischemia are required to produce a similar effect on heart of Goto-Kakizaki rats (Tsang et al., 2005). This study also demonstrated that the ischemic preconditioning stimuli should be strong enough to increase the Akt phosphorylation necessary to achieve threshold for cardioprotection. Based on this observation, in a follow-up study this group of investigators evaluated the effect of glimepiride -- a known activator of Akt. They observed that glimepiride treatment was able to lower the threshold for ischemic preconditioning in such a manner that both 1 and 3 cycles of ischemic preconditioning involving cycle(s) of 5 minutes of global ischemia and 10 minutes of reperfusion produced a cardioprotective effect in diabetic heart isolated from Goto-Kakizaki rats (Hausenloy et al., 2013). This indicates that while a higher intensity of preconditioning stimuli is required to observe protective effects on ischemic myocardium, the treatment with glimepiride, an anti-diabetic drug, lowers the threshold of preconditioning stimuli required to observe the protective effect of ischemic preconditioning on diabetic heart. This study also demonstrates that glimepiride reverses the effect of diabetes on ischemic preconditioning, possibly due to its interaction with the pathways mediating the

beneficial effect of ischemic preconditioning on myocardium. However, further studies with other drugs that activate conditioning pathways are required to further confirm this premise.

A limited number of studies have evaluated mechanisms involved in suppression of conditioning-induced ischemia tolerance in diabetic animals. The inhibitory effect of diabetes on myocardial preconditioning is implicated by dysfunction in sarcolemmal  $K_{ATP}$  channels (del Valle et al., 2003). Dysfunction of mitochondrial  $K_{ATP}$  channels is also proposed to mediate diabetic attenuation of the protective effect of myocardial preconditioning (Ghosh et al., 2001). Glycogen synthase kinase-3 $\beta$  also mediated diabetic attenuation of the beneficial effect of ischemic preconditioning (4 cycles of 5-min ischemia and 5-min reperfusion) on the isolated rat heart exposed to ischemia-reperfusion injury (Yadav et al., 2010). However, better understanding of conditioning mechanisms affected in diabetes may help design novel strategies to induce ischemia tolerance in diabetics.

The effect of ischemic preconditioning has also been evaluated in other tissues and organs in diabetes. Ischemic preconditioning 72 h prior to ischemia exerts a significant ameliorative effect on transient focal cerebral ischemia-induced brain injury in both non-diabetic and diabetic rats (seven days post-streptozotocin injection) with a concomitant upregulation of pro-survival miRNAs in the infarcted brain area (Altintas et al., 2016a; Altintas et al., 2016b). It is possible that longer (several weeks) and not short duration (7 days) of diabetes may attenuate the protective effect of ischemic preconditioning. Hypoxia/hypoglycemia-induced preconditioning decreases delayed ischemic cell death and the extent of loss of functional electrical activity in organotypic slice cultures (Badaut et al., 2005). Previously, our laboratory showed that prior exposure to recurrent hypoglycemia enhances ischemic brain injury in insulin-treated diabetic rats (Dave et al., 2011b) and oxygen glucose deprivation-induced damage in hippocampal organotypic slices (Dave et al., 2011a). Ischemic conditioning induced by weekly repetitions of 5 min of retinal ischemia in streptozotocin diabetic rats prevented axoglial changes in the optic pathway. These axoglial changes include deficits in the anterograde transport from the retina to the superior colliculus, increase in astrocyte reactivity, ultrastructural alteration in myelin, and altered oligodendrocyte morphology in the distal portion of the optic nerve (Fernandez et al., 2011; Fernandez et al., 2012). These studies again emphasize that stronger preconditioning stimuli (weekly repetitions of conditioning stimuli) may be needed to induce ischemia tolerance in diabetic eyes of diabetic individuals. Ischemic preconditioning of intestine and liver of diabetic rats prior to severe ischemia in the respective organs reduced leukocyte infiltration in the lungs (Thomaz Neto et al., 2013). In summary, a longer duration of diabetes may attenuate the protective effects of ischemic preconditioning, and stronger conditioning stimuli may be required to induce ischemia tolerance in diabetic animals. However, further detailed studies are required to understand the effect of chronic diabetes on ischemia-reperfusion injury in brain, kidney, intestines and liver.

### Ischemic postconditioning and protection in diabetes

Ischemic postconditioning exerts a potent protective effect on tissues or organs subjected to otherwise severe ischemic insult (Zhao et al., 2003; Kin et al., 2004; Kin et al., 2005; Zhao et al., 2006). Details appear in earlier reviews (Zhao, 2007, 2009; Zhao et al., 2012; Theodoraki et al., 2016). Clinical studies also confirmed the beneficial effects of ischemic postconditioning on ischemic myocardium (Staat et al., 2005; Hansen et al., 2010). However, ischemic postconditioning induced by three or six 10-s cycles of reperfusion-reocclusion on an isolated mouse

heart preparation did not produce any beneficial effect on the extent of infarct size in the *db/db* mouse model of T2D and streptozotocin-induced mouse model of T1D (Przyklenk et al., 2011). Four to five weeks of streptozotocin-induced diabetes attenuates the ameliorative effect of ischemic postconditioning on ischemia-reperfusion injury-induced myocardial infarction in rats *in vivo* (Drenger et al., 2011). Similarly, the ischemic postconditioning-induced reduction in myocardial infarct size (observed in control mice) was not observed in *db/db* mice and leptin-deficient obese (*ob/ob*) mice subjected to myocardial ischemia (Bouhidel et al., 2008; Zhu et al., 2012). These studies indicate that ischemic postconditioning does not confer beneficial effects on ischemia myocardium in multiple models of diabetes. The effect of ischemic postconditioning on other ischemic tissues and organs in diabetic animals remains to be established.

### Remote ischemic conditioning and protection in diabetes

Preclinical data demonstrates that remote ischemic conditioning (RIC) can induce protection against ischemic damage (Hausenloy and Yellon, 2008; Heusch et al., 2015; Pickard et al., 2015; Sivaraman et al., 2015; Giannopoulos et al., 2017). Remote ischemic preconditioning elicited by transient liver ischemia reduces the incidence of ventricular tachyarrhythmias in both non-diabetic and diabetic rats. However, in comparison to non-diabetic rats, remote preconditioning failed to exert a beneficial effect on atrioventricular block in diabetic rats (Hu et al., 2017).

Acute hyperglycemia attenuates the cardioprotective effect of remote femoral artery and vein occlusion-induced remote preconditioning on ischemic myocardium (induced by transient occlusion of the left anterior descending coronary artery for 40 min) *in vivo* (Baranyai et al., 2015). This study demonstrated that remote preconditioning in acute hyperglycemic rats did not influence infarct size but increased incidence as well as duration of arrhythmias. They also hypothesized that deterioration in cardioprotection by remote preconditioning in acute hyperglycemic rats may be due to increased nitrate stress. Remote ischemic preconditioning induced by bilateral femoral artery occlusion exerts a significant protective effect on left coronary artery occlusion-induced myocardial ischemia and associated infarct size in nondiabetic rats. However, this protective effect was absent in diabetic rats (Kiss et al., 2014).

A recent double blinded, randomized, placebo-controlled, multicenter clinical study demonstrated that remote ischemic preconditioning induced by four cycles of intermittent upper arm ischemia-reperfusion exerts a protective effect on contrast-induced nephropathy in non-diabetic subjects with moderate renal dysfunction undergoing percutaneous coronary intervention (Moretti et al., 2018). However, they did not observe a similar protective effect of remote ischemic preconditioning (RIC) on diabetic patients (Moretti et al., 2018). Clinical studies evaluating the efficacy of RIC on the ischemic myocardium (in terms of levels of marker of ischemic cell death) showed either beneficial (Cheung et al., 2006; Hausenloy et al., 2007; Ali et al., 2010; Kottenberg et al., 2012; Candilio et al., 2015) or no effect (Gunaydin et al., 2000; Karuppsamy et al., 2011; Lucchinetti et al., 2012; McCrindle et al., 2014). It is plausible that such a mixed result may be due to patient populations with various comorbidities such as diabetes. Overall, studies evaluating the effect of diabetes on remote conditioning are minimal, and more studies are required to understand the effect of diabetes on the protective effect of remote ischemic preconditioning.

### Pharmacological conditioning and protection in diabetes

Activating ischemia tolerance pathways via pharmacological conditioning exerts a protective effect against ischemic

damage (Julier et al., 2003; Luca et al., 2011; Wang et al., 2011; Shi et al., 2013). However, the effect of pharmacological conditioning on ischemic damage in diabetic animals or human subjects is not well understood. Acute hyperglycemia attenuates isoflurane postconditioning-induced reduction in myocardial infarct size and creatine kinase levels in a rabbit heart ischemia-reperfusion model (Raphael et al., 2010). In addition, moderate to severe hyperglycemia prevented the cardioprotective effect of isoflurane preconditioning on dogs subjected to left anterior descending coronary artery occlusion-reperfusion-induced injury (Kehl et al., 2002). The effect of isoflurane-induced preconditioning of the myocardium is also attenuated in alloxan- and streptozotocin-treated dogs (Tanaka et al., 2002). Sevoflurane postconditioning-induced cardioprotection is lost in streptozotocin-diabetic rats (Drenger et al., 2011; Tai et al., 2012). Erythropoietin postconditioning-induced reduction in myocardial infarct size was observed in a cohort of rats with high-fat diet-induced insulin resistance syndrome. However, the protection was abrogated in streptozotocin-induced diabetic rats (Ghaboura et al., 2011). This study also concluded that disruption of glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ) signaling may be responsible for observed abolished erythropoietin-induced cardioprotection in streptozotocin-induced diabetic rats. The cardioprotective effect of erythropoietin and [D-Ala<sup>2</sup>, D-Leu<sup>5</sup>]-enkephalin acetate (a delta-opioid receptor agonist) preconditioning was abolished in a rat model of T2D (Otsuka-Long-Evans-Tokushima fatty rats) (Hotta et al., 2010). However, blockade of angiotensin II type 1 receptor for a period of 2 weeks restored the cardioprotective effect of erythropoietin preconditioning in diabetic animals (Hotta et al., 2010). Similarly, morphine preconditioning did not affect ischemia-reperfusion-induced infarct size in streptozotocin-diabetic rats, but reduced infarct size in the nondiabetic group (Gross et al., 2007). This study also concluded that this abolished morphine-induced cardioprotection in diabetic rats was due to reduced activation of mediators of GSK-3 $\beta$  signaling. While isoflurane-induced preconditioning of heart produced a beneficial effect in non-diabetic Wistar rats, it failed to exert a similar effect on Goto-Kakizaki rats (Matsumoto et al., 2009). However, olprinone preconditioning protected hearts of Goto-Kakizaki rats against myocardial infarction potentially via the phosphatidylinositol 3-kinase-Akt pathway (Matsumoto et al., 2009). Thus, the literature demonstrates that the presence of diabetes reduces the ameliorative effect of pharmacological preconditioning on ischemic tissue. However, more research is required to understand the mechanistic basis of the effect of diabetes on pharmacological conditioning to induce ischemia tolerance.

### Antidiabetic therapy and effects of conditioning on ischemia-reperfusion injury

There are multiple therapeutic approaches proposed to lower cardiovascular risk among diabetics. One of the approaches involves pharmacological control of blood glucose levels using drugs such as metformin, sulfonylureas, PPAR- $\gamma$  agonists (thiazolidinedione),  $\alpha$ -glucosidase inhibitors and insulin. Other approaches include antihypertensive drugs including ACE-inhibitors, angiotensin-2 receptor blockers, lifestyle interventions, anti-hyperlipidemic drugs, and anti-platelet agents (International Diabetes Federation, 2012). However, given the widespread prevalence of diabetes and associated increases in cardiovascular diseases worldwide, it is also important to understand the potential interactions between these therapeutic approaches and the promising beneficial effect of conditioning. A clinical study by Cleveland et al. evaluated the effect of long-term exposure to oral anti-hyperglycemic agents vs. insulin therapy on ischemic preconditioning in protecting isolated myocardium (right atrial trabeculae) against

ischemic damage (Cleveland et al., 1997). They observed that ischemic preconditioning was able to induce greater recovery of developed force in myocardium from control and in diabetic patients receiving long-term insulin treatment. However, ischemic preconditioning was not able to induce recovery of developed force in myocardium from diabetic patients receiving long-term oral hypoglycemic agents. Glibenclamide and glimepiride are sulfonylurea drugs used for the treatment of T2D (Langtry and Balfour, 1998; Korytkowski et al., 2002). The mechanism of action of their anti-diabetic effects involves inhibition of  $K_{ATP}$  channels in pancreatic  $\beta$ -cells (Kramer et al., 1995; Ueda et al., 1999), which are key regulators of insulin release via enhanced intracellular calcium influx (Miki et al., 1998). Glibenclamide attenuates ischemic preconditioning in various laboratory models (Schulz et al., 1994; Miura et al., 1995; Hoag et al., 1997) as well as in human subjects during brief repeated coronary occlusions (in terms of ST segment shift and pain severity) (Tomai et al., 1994). In addition, glibenclamide also attenuates the exercise-induced beneficial effect of the warm-up phenomenon in diabetic subjects (Ovunc, 2000). A double-blind placebo-controlled clinical study showed that while glibenclamide attenuates preconditioning, glimepiride, another  $K_{ATP}$  channel inhibitor, maintains the effects of preconditioning in myocardium (Klepzig et al., 1999). It is plausible that glibenclamide and glimepiride may affect other pathways of ischemia tolerance differently, which may explain their differential effects on ischemia tolerance in myocardium.

Metformin belongs to the biguanide category of orally administered anti-diabetic drugs and is the first-line drug for the treatment of T2D (Inzucchi et al., 2012). It acts by reducing "hepatic glucose output" and "increasing insulin-mediated glucose disposal" (Bailey and Turner, 1996), and is one of the most widely used drugs for treatment of T2D (Inzucchi et al., 2012). Metformin, when administered at the time of reperfusion, reduces myocardial infarct size in both the non-diabetic (Wistar rats) and diabetic (Goto-Kakizaki rats) heart, possibly via activation of phosphoinositide 3-kinase, and is associated with Akt phosphorylation (Bhamra et al., 2008). In addition, metformin administration before ischemia or at the time of reperfusion decreases myocardial injury in both nondiabetic and diabetic mice (db/db) (Calvert et al., 2008). Pretreatment with metformin 24 hours prior to permanent middle cerebral artery occlusion reduces infarct size, neurological deficits, and cell apoptosis in ischemic brain (Jiang et al., 2014). Continuous treatment with metformin daily for a period of 3 weeks induces ischemia tolerance resulting in reduced focal cerebral ischemia-induced infarct and neurological deficits, potentially by suppressing NF- $\kappa$ B-mediated inflammatory pathways (Zhu et al., 2015).

There are many new and established anti-diabetic drugs tested for their effect on ischemia tolerance in diabetic animals. 5-hydroxydecanoate, a  $K_{ATP}$  channel inhibitor, abrogates the beneficial effect of heat stress preconditioning on heart (Hoag et al., 1997). Repaglinide is reported to attenuate protective effects of myocardial preconditioning in T2D subjects (Rahmi et al., 2013). A study aimed to determine the effect of various pharmacological preconditioning agents on myocardial ischemia using human diabetic myocardium observed that phenylephrine, adenosine, or diazoxide failed to protect diabetic myocardium. In contrast, phorbol-12-myristate-13-acetate (protein kinase C activator), or anisomycin (p38 mitogen-activated protein kinase activator) induced a significant ischemia tolerance in diabetic myocardium (Hassouna et al., 2006). WY-14643, a peroxisome proliferator-activated receptor- $\alpha$  agonist, exerts a significant cardioprotective effect on ischemic myocardium in control and Goto-Kakizaki rats, possibly via the activation of phosphoinositide 3-kinase/Akt and nitric oxide pathways

(Bulhak et al., 2009). Similarly, rosiglitazone treatment in Zucker diabetic fatty rats exerted a marked cardioprotective effect on myocardium subjected to ischemia (Yue et al., 2005). These studies suggest that the effect of diabetes on ischemia tolerance may vary depending on which glucose-lowering therapy is given. This mixed effect of anti-diabetic drugs on ischemia-reperfusion injury during diabetes also identifies a need for more research in this area.

### Future Directions

Overall, the studies discussed in this article clearly demonstrate that diabetes as a comorbid condition affects the beneficial effect of conditioning on ischemic injury. However, studies ascertaining the potential effect of chronic diabetes on the protection afforded by conditioning on ischemic injury in organs like brain, kidney, liver, skeletal muscles and intestines are broadly missing. Given the number of metabolic pathways altered during diabetes and variability in the animal models used to study the effect of diabetes on conditioning, confirmatory studies determining the effect of diabetes on the ameliorative potential of different forms of conditioning would help better characterize the effect of diabetes on the induction of ischemia tolerance. In addition, studies aimed at better understanding the mechanisms by which diabetes affects the beneficial effect of conditioning are also needed. It is also important to evaluate the protective effect of conditioning in treated diabetes. Furthermore, most of the available studies have evaluated the effect of diabetes on ischemia tolerance in young animals. The Centers for Disease Control and Prevention estimates that the prevalence of diabetes is more than 25% in those aged 65 years or older (Centers for Disease Control and Prevention, 2017). The risk of mortality and morbidity due to cardiovascular diseases greatly increases with advancing age in diabetic subjects, as reviewed previously (Cigolle et al., 2009; Kirkman et al., 2012). Therefore, it is important to evaluate the effect of diabetes on conditioning in aged animals. The Stroke Therapy Academic Industry Roundtable (STAIR) research recommendations highlight the clinical significance of studying the effect of comorbid conditions like diabetes on ischemic injury in brain. Application of STAIR criteria for assessing the effect of different forms of conditioning on ischemic tissue would help increase translational potential of various ischemia tolerance induction paradigms.

### Summary

Most of the studies aimed at understanding the effect of diabetes on different forms of conditioning demonstrate that diabetes attenuates the effects of various conditioning paradigms, potentially via affecting ischemic tolerance induction pathways. Detailed characterization of mechanisms behind diabetic attenuation of conditioning may help better tailor various conditioning paradigms to induce ischemia tolerance in diabetic patients.

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### References

- Adstamongkonkul D, Hess, D.C., (2017) Ischemic Conditioning and neonatal hypoxic ischemic encephalopathy: a literature review. *Conditioning Medicine* 1:9-16.
- Afkarian M, Zelnick LR, Hall YN, Heagerty PJ, Tuttle K, Weiss NS, de Boer IH (2016) Clinical Manifestations of Kidney Disease Among US Adults With Diabetes, 1988-2014. *JAMA* 316:602-610.
- Alegria JR, Miller TD, Gibbons RJ, Yi QL, Yusuf S,

- Collaborative Organization of RheothRx Evaluation Trial I (2007) Infarct size, ejection fraction, and mortality in diabetic patients with acute myocardial infarction treated with thrombolytic therapy. *Am Heart J* 154:743-750.
- Ali N, Rizwi F, Iqbal A, Rashid A (2010) Induced remote ischemic preconditioning on ischemia-reperfusion injury in patients undergoing coronary artery bypass. *J Coll Physicians Surg Pak* 20:427-431.
- Almdal T, Scharling H, Jensen JS, Vestergaard H (2004) The independent effect of type 2 diabetes mellitus on ischemic heart disease, stroke, and death: a population-based study of 13,000 men and women with 20 years of follow-up. *Arch Intern Med* 164:1422-1426.
- Altintas O, Kumas M, Altintas MO (2016a) Neuroprotective effect of ischemic preconditioning via modulating the expression of adropin and oxidative markers against transient cerebral ischemia in diabetic rats. *Peptides* 79:31-38.
- Altintas O, Ozgen Altintas M, Kumas M, Asil T (2016b) Neuroprotective effect of ischemic preconditioning via modulating the expression of cerebral miRNAs against transient cerebral ischemia in diabetic rats. *Neurol Res* 38:1003-1011.
- American Diabetes Association (2014) Diagnosis and classification of diabetes mellitus. *Diabetes Care* 37 Suppl 1:S81-90.
- American Optometric Association (2018) Diabetic Retinopathy. In: <https://www.aoa.org/patients-and-public/eye-and-vision-problems/glossary-of-eye-and-vision-conditions/diabetic-retinopathy>.
- Badaut J, Hirt L, Price M, de Castro Ribeiro M, Magistretti PJ, Regli L (2005) Hypoxia/hypoglycemia preconditioning prevents the loss of functional electrical activity in organotypic slice cultures. *Brain Res* 1051:117-122.
- Bailey CJ, Turner RC (1996) Metformin. *N Engl J Med* 334:574-579.
- Baranyai T, Nagy CT, Koncsos G, Onodi Z, Karolyi-Szabo M, Makkos A, Varga ZV, Ferdinandy P, Giricz Z (2015) Acute hyperglycemia abolishes cardioprotection by remote ischemic preconditioning. *Cardiovasc Diabetol* 14:151.
- Bhalla V, Zhao B, Azar KM, Wang EJ, Choi S, Wong EC, Fortmann SP, Palaniappan LP (2013) Racial/ethnic differences in the prevalence of proteinuric and nonproteinuric diabetic kidney disease. *Diabetes Care* 36:1215-1221.
- Bhamra GS, Hausenloy DJ, Davidson SM, Carr RD, Paiva M, Wynne AM, Mocanu MM, Yellon DM (2008) Metformin protects the ischemic heart by the Akt-mediated inhibition of mitochondrial permeability transition pore opening. *Basic Res Cardiol* 103:274-284.
- Bouchard JF, Lamontagne D (1998) Protection afforded by preconditioning to the diabetic heart against ischaemic injury. *Cardiovasc Res* 37:82-90.
- Bouhidel O, Pons S, Souktani R, Zini R, Berdeaux A, Ghaleh B (2008) Myocardial ischemic postconditioning against ischemia-reperfusion is impaired in ob/ob mice. *Am J Physiol Heart Circ Physiol* 295:H1580-1586.
- Bulhak AA, Jung C, Ostenson CG, Lundberg JO, Sjoquist PO, Pernow J (2009) PPAR-alpha activation protects the type 2 diabetic myocardium against ischemia-reperfusion injury: involvement of the PI3-Kinase/Akt and NO pathway. *Am J Physiol Heart Circ Physiol* 296:H719-727.
- Calvert JW, Gundewar S, Jha S, Greer JJ, Bestermann WH, Tian R, Lefer DJ (2008) Acute metformin therapy confers cardioprotection against myocardial infarction via AMPK-eNOS-mediated signaling. *Diabetes* 57:696-705.
- Candilio L, Malik A, Ariti C, Barnard M, Di Salvo C, Lawrence D, Hayward M, Yap J, Roberts N, Sheikh A, Kolvekar S, Hausenloy DJ, Yellon DM (2015) Effect of remote ischaemic preconditioning on clinical outcomes in patients undergoing cardiac bypass surgery: a randomised controlled clinical trial. *Heart* 101:185-192.
- Centers for Disease Control and Prevention (2003) Self-reported heart disease and stroke among adults with and without diabetes--United States, 1999-2001. *MMWR Morb Mortal Wkly Rep* 52:1065-1070.
- Centers for Disease Control and Prevention (2017) National Diabetes Statistics Report. U.S. Dept of Health and Human Services. In: <https://www.cdc.gov/diabetes/pdfs/data/statistics/national-diabetes-statistics-report.pdf>.
- Cheung MM, Kharbanda RK, Konstantinov IE, Shimizu M, Frndova H, Li J, Holtby HM, Cox PN, Smallhorn JF, Van Arsdell GS, Redington AN (2006) Randomized controlled trial of the effects of remote ischemic preconditioning on children undergoing cardiac surgery: first clinical application in humans. *J Am Coll Cardiol* 47:2277-2282.
- Cigolle CT, Blaum CS, Halter JB (2009) Diabetes and cardiovascular disease prevention in older adults. *Clin Geriatr Med* 25:607-641, vii-viii.
- Cipollone F, Iezzi A, Fazio M, Zucchelli M, Pini B, Cuccurullo C, De Cesare D, De Blasis G, Muraro R, Bei R, Chiarelli F, Schmidt AM, Cuccurullo F, Mezzetti A (2003) The receptor RAGE as a progression factor amplifying arachidonate-dependent inflammatory and proteolytic response in human atherosclerotic plaques: role of glycemic control. *Circulation* 108:1070-1077.
- Cleveland JC, Jr., Meldrum DR, Cain BS, Banerjee A, Harken AH (1997) Oral sulfonylurea hypoglycemic agents prevent ischemic preconditioning in human myocardium. Two paradoxes revisited. *Circulation* 96:29-32.
- Dave KR, Pileggi A, Raval AP (2011a) Recurrent hypoglycemia increases oxygen glucose deprivation-induced damage in hippocampal organotypic slices. *Neurosci Lett* 496:25-29.
- Dave KR, Saul I, Prado R, Busto R, Perez-Pinzon MA (2006) Remote organ ischemic preconditioning protect brain from ischemic damage following asphyxial cardiac arrest. *Neurosci Lett* 404:170-175.
- Dave KR, Saul I, Busto R, Ginsberg MD, Sick TJ, Perez-Pinzon MA (2001) Ischemic preconditioning preserves mitochondrial function after global cerebral ischemia in rat hippocampus. *J Cereb Blood Flow Metab* 21:1401-1410.
- Dave KR, Lange-Asschenfeldt C, Raval AP, Prado R, Busto R, Saul I, Perez-Pinzon MA (2005) Ischemic preconditioning ameliorates excitotoxicity by shifting glutamate/gamma-aminobutyric acid release and biosynthesis. *J Neurosci Res* 82:665-673.
- Dave KR, Tamariz J, Desai KM, Brand FJ, Liu A, Saul I, Bhattacharya SK, Pileggi A (2011b) Recurrent hypoglycemia exacerbates cerebral ischemic damage in streptozotocin-induced diabetic rats. *Stroke* 42:1404-1411.
- de Ferranti SD, de Boer IH, Fonseca V, Fox CS, Golden SH, Lavie CJ, Magge SN, Marx N, McGuire DK, Orchard TJ, Zinman B, Eckel RH (2014) Type 1 diabetes mellitus and cardiovascular disease: a scientific statement from the American Heart Association and American Diabetes Association. *Circulation* 130:1110-1130.
- del Valle HF, Lascano EC, Negroni JA, Crottogini AJ (2003) Absence of ischemic preconditioning protection in diabetic sheep hearts: role of sarcolemmal KATP channel dysfunction. *Mol Cell Biochem* 249:21-30.
- Drenger B, Ostrovsky IA, Barak M, Nechemia-Arbely Y, Ziv E, Axelrod JH (2011) Diabetes blockade of sevoflurane postconditioning is not restored by insulin in the rat heart: phosphorylated signal transducer and activator of transcription 3- and phosphatidylinositol 3-kinase-

- mediated inhibition. *Anesthesiology* 114:1364-1372.
- Ebel D, Mullenheim J, Frassdorf J, Heinen A, Huhn R, Bohlen T, Ferrari J, Sudkamp H, Preckel B, Schlack W, Thamer V (2003) Effect of acute hyperglycaemia and diabetes mellitus with and without short-term insulin treatment on myocardial ischaemic late preconditioning in the rabbit heart in vivo. *Pflugers Arch* 446:175-182.
- Emerging Risk Factors Collaboration, Sarwar N, Gao P, Seshasai SR, Gobin R, Kaptoge S, Di Angelantonio E, Ingelsson E, Lawlor DA, Selvin E, Stampfer M, Stehouwer CD, Lewington S, Pennells L, Thompson A, Sattar N, White IR, Ray KK, Danesh J (2010) Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. *Lancet* 375:2215-2222.
- Fernandez DC, Sande PH, Chianelli MS, Aldana Marcos HJ, Rosenstein RE (2011) Induction of ischemic tolerance protects the retina from diabetic retinopathy. *Am J Pathol* 178:2264-2274.
- Fernandez DC, Pasquini LA, Dorfman D, Aldana Marcos HJ, Rosenstein RE (2012) Ischemic conditioning protects from axoglial alterations of the optic pathway induced by experimental diabetes in rats. *PLoS One* 7:e51966.
- Fonseca V, Desouza C, Asnani S, Jialal I (2004) Nontraditional risk factors for cardiovascular disease in diabetes. *Endocr Rev* 25:153-175.
- Fox CS GS, Anderson C, Bray GA, Burke LE, de Boer IH, Deedwania P, Eckel RH, Ershow AG, Fradkin J, Inzucchi SE, Kosiborod M, Nelson RG, Patel MJ, Pignone M, Quinn L, Schauer PR, Selvin E, Vafiadis DK; American Heart Association Diabetes Committee of the Council on Lifestyle and Cardiometabolic Health; Council on Clinical Cardiology, Council on Cardiovascular and Stroke Nursing, Council on Cardiovascular Surgery and Anesthesia, Council on Quality of Care and Outcomes Research; American Diabetes Association (2015) Update on Prevention of Cardiovascular Disease in Adults With Type 2 Diabetes Mellitus in Light of Recent Evidence: A Scientific Statement From the American Heart Association and the American Diabetes Association. *Circulation* 132:691-718.
- Fuller JH, Stevens LK, Wang SL (2001) Risk factors for cardiovascular mortality and morbidity: the WHO Multinational Study of Vascular Disease in Diabetes. *Diabetologia* 44 Suppl 2:S54-64.
- Ghaboura N, Tamareille S, Ducluzeau PH, Grimaud L, Loufrani L, Croue A, Tourmen Y, Henrion D, Furber A, Prunier F (2011) Diabetes mellitus abrogates erythropoietin-induced cardioprotection against ischemic-reperfusion injury by alteration of the RISK/GSK-3 $\beta$  signaling. *Basic Res Cardiol* 106:147-162.
- Ghosh S, Standen NB, Galinanes M (2001) Failure to precondition pathological human myocardium. *J Am Coll Cardiol* 37:711-718.
- Giannopoulos G, Vrachatis DA, Panagopoulou V, Vavuranakis M, Cleman MW, Deftereos S (2017) Remote Ischemic Conditioning and Renal Protection. *J Cardiovasc Pharmacol Ther* 22:321-329.
- Gross ER, Hsu AK, Gross GJ (2007) Diabetes abolishes morphine-induced cardioprotection via multiple pathways upstream of glycogen synthase kinase-3 $\beta$ . *Diabetes* 56:127-136.
- Gunaydin B, Cakici I, Soncul H, Kalaycioglu S, Cevik C, Sancak B, Kanzik I, Karadenizli Y (2000) Does remote organ ischaemia trigger cardiac preconditioning during coronary artery surgery? *Pharmacol Res* 41:493-496.
- Hansen PR, Thibault H, Abdulla J (2010) Postconditioning during primary percutaneous coronary intervention: a review and meta-analysis. *Int J Cardiol* 144:22-25.
- Hassouna A, Loubani M, Matata BM, Fowler A, Standen NB, Galinanes M (2006) Mitochondrial dysfunction as the cause of the failure to precondition the diabetic human myocardium. *Cardiovasc Res* 69:450-458.
- Hausenloy DJ, Yellon DM (2008) Remote ischaemic preconditioning: underlying mechanisms and clinical application. *Cardiovasc Res* 79:377-386.
- Hausenloy DJ, Wynne AM, Mocanu MM, Yellon DM (2013) Glimepiride treatment facilitates ischemic preconditioning in the diabetic heart. *J Cardiovasc Pharmacol Ther* 18:263-269.
- Hausenloy DJ, Mwamure PK, Venugopal V, Harris J, Barnard M, Grundy E, Ashley E, Vichare S, Di Salvo C, Kolvekar S, Hayward M, Keogh B, MacAllister RJ, Yellon DM (2007) Effect of remote ischaemic preconditioning on myocardial injury in patients undergoing coronary artery bypass graft surgery: a randomised controlled trial. *Lancet* 370:575-579.
- Hausenloy DJ et al. (2016) Ischaemic conditioning and targeting reperfusion injury: a 30 year voyage of discovery. *Basic Res Cardiol* 111:70.
- Heusch G, Botker HE, Przyklenk K, Redington A, Yellon D (2015) Remote ischemic conditioning. *J Am Coll Cardiol* 65:177-195.
- Hoag JB, Qian YZ, Nayeem MA, D'Angelo M, Kukreja RC (1997) ATP-sensitive potassium channel mediates delayed ischemic protection by heat stress in rabbit heart. *Am J Physiol* 273:H2458-2464.
- Hotta H, Miura T, Miki T, Togashi N, Maeda T, Kim SJ, Tanno M, Yano T, Kuno A, Itoh T, Satoh T, Terashima Y, Ishikawa S, Shimamoto K (2010) Angiotensin II type 1 receptor-mediated upregulation of calcineurin activity underlies impairment of cardioprotective signaling in diabetic hearts. *Circ Res* 106:129-132.
- Hu Z, Chen M, Zhang P, Liu J, Abbott GW (2017) Remote ischemic preconditioning differentially attenuates post-ischemic cardiac arrhythmia in streptozotocin-induced diabetic versus nondiabetic rats. *Cardiovasc Diabetol* 16:57.
- Huo X GL, Guo L, Xu W, Wang W, Zhi X, Li L, Ren Y, Qi X, Sun Z, Li W, Ji Q, Ran X, Su B, Hao C, Lu J, Guo X, Zhuo H, Zhang D, Pan C, Weng J, Hu D, Yang X, Ji L (2016) Risk of non-fatal cardiovascular diseases in early-onset versus late-onset type 2 diabetes in China: a cross-sectional study. *Lancet Diabetes Endocrinol* 4:115-124.
- International Diabetes Federation (2012) Global Guideline for Type 2 Diabetes, Brussels <https://www.idf.org/e-library/guidelines/79-global-guideline-for-type-2-diabetes>. In.
- Inzucchi SE, Bergenstal RM, Buse JB, Diamant M, Ferrannini E, Nauck M, Peters AL, Tsapas A, Wender R, Matthews DR (2012) Management of hyperglycaemia in type 2 diabetes: a patient-centered approach. Position statement of the American Diabetes Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetologia* 55:1577-1596.
- Janghorbani M, Hu FB, Willett WC, Li TY, Manson JE, Logroscino G, Rexrode KM (2007) Prospective study of type 1 and type 2 diabetes and risk of stroke subtypes: the Nurses' Health Study. *Diabetes Care* 30:1730-1735.
- Jia Z, Lian W, Shi H, Cao C, Han S, Wang K, Li M, Zhang X (2017) Ischemic Postconditioning Protects Against Intestinal Ischemia/Reperfusion Injury via the HIF-1 $\alpha$ /miR-21 Axis. *Sci Rep* 7:16190.
- Jiang T, Yu JT, Zhu XC, Wang HF, Tan MS, Cao L, Zhang QQ, Gao L, Shi JQ, Zhang YD, Tan L (2014) Acute metformin preconditioning confers neuroprotection against focal cerebral ischaemia by pre-activation of AMPK-dependent



- autophagy. *Br J Pharmacol* 171:3146-3157.
- Jonker SJ, Menting TP, Warle MC, Ritskes-Hoitinga M, Wever KE (2016) Preclinical Evidence for the Efficacy of Ischemic Postconditioning against Renal Ischemia-Reperfusion Injury, a Systematic Review and Meta-Analysis. *PLoS One* 11:e0150863.
- Julier K, da Silva R, Garcia C, Bestmann L, Frascarolo P, Zollinger A, Chassot PG, Schmid ER, Turina MI, von Segesser LK, Pasch T, Spahn DR, Zaugg M (2003) Preconditioning by sevoflurane decreases biochemical markers for myocardial and renal dysfunction in coronary artery bypass graft surgery: a double-blinded, placebo-controlled, multicenter study. *Anesthesiology* 98:1315-1327.
- Karuppasamy P, Chaubey S, Dew T, Musto R, Sherwood R, Desai J, John L, Shah AM, Marber MS, Kunst G (2011) Remote intermittent ischemia before coronary artery bypass graft surgery: a strategy to reduce injury and inflammation? *Basic Res Cardiol* 106:511-519.
- Katakam PV, Jordan JE, Snipes JA, Tulbert CD, Miller AW, Busija DW (2007) Myocardial preconditioning against ischemia-reperfusion injury is abolished in Zucker obese rats with insulin resistance. *Am J Physiol Regul Integr Comp Physiol* 292:R920-926.
- Kehl F, Krolkowski JG, Mraovic B, Pagel PS, Warltier DC, Kersten JR (2002) Hyperglycemia prevents isoflurane-induced preconditioning against myocardial infarction. *Anesthesiology* 96:183-188.
- Kersten JR, Schmeling TJ, Orth KG, Pagel PS, Warltier DC (1998) Acute hyperglycemia abolishes ischemic preconditioning in vivo. *Am J Physiol* 275:H721-725.
- Kersten JR, Toller WG, Gross ER, Pagel PS, Warltier DC (2000) Diabetes abolishes ischemic preconditioning: role of glucose, insulin, and osmolality. *Am J Physiol Heart Circ Physiol* 278:H1218-1224.
- Kin H, Zhao ZQ, Sun HY, Wang NP, Corvera JS, Halkos ME, Kerendi F, Guyton RA, Vinten-Johansen J (2004) Postconditioning attenuates myocardial ischemia-reperfusion injury by inhibiting events in the early minutes of reperfusion. *Cardiovasc Res* 62:74-85.
- Kin H, Zatta AJ, Lofye MT, Amerson BS, Halkos ME, Kerendi F, Zhao ZQ, Guyton RA, Headrick JP, Vinten-Johansen J (2005) Postconditioning reduces infarct size via adenosine receptor activation by endogenous adenosine. *Cardiovasc Res* 67:124-133.
- Kirkman MS, Briscoe VJ, Clark N, Florez H, Haas LB, Halter JB, Huang ES, Korytkowski MT, Munshi MN, Odegaard PS, Pratley RE, Swift CS (2012) Diabetes in older adults. *Diabetes Care* 35:2650-2664.
- Kiss A, Tratsiakovich Y, Gonon AT, Fedotovskaya O, Lanner JT, Andersson DC, Yang J, Pernow J (2014) The role of arginase and rho kinase in cardioprotection from remote ischemic preconditioning in non-diabetic and diabetic rat in vivo. *PLoS One* 9:e104731.
- Kissela BM, Khoury J, Kleindorfer D, Woo D, Schneider A, Alwell K, Miller R, Ewing I, Moomaw CJ, Szaflarski JP, Gebel J, Shukla R, Broderick JP (2005) Epidemiology of ischemic stroke in patients with diabetes: the greater Cincinnati/Northern Kentucky Stroke Study. *Diabetes Care* 28:355-359.
- Kitagawa K, Matsumoto M, Tagaya M, Hata R, Ueda H, Niinobe M, Handa N, Fukunaga R, Kimura K, Mikoshiba K, et al. (1990) 'Ischemic tolerance' phenomenon found in the brain. *Brain Res* 528:21-24.
- Klepzig H, Kober G, Matter C, Luus H, Schneider H, Boedeker KH, Kiowski W, Amann FW, Gruber D, Harris S, Burger W (1999) Sulfonylureas and ischaemic preconditioning; a double-blind, placebo-controlled evaluation of glimepiride and glibenclamide. *Eur Heart J* 20:439-446.
- Kobayashi S, Harris VA, Welsh FA (1995) Spreading depression induces tolerance of cortical neurons to ischemia in rat brain. *J Cereb Blood Flow Metab* 15:721-727.
- Kolber MR, Scrimshaw C (2014) Family history of cardiovascular disease. *Can Fam Physician* 60:1016.
- Koronowski KB, Dave KR, Saul I, Camarena V, Thompson JW, Neumann JT, Young JI, Perez-Pinzon MA (2015) Resveratrol Preconditioning Induces a Novel Extended Window of Ischemic Tolerance in the Mouse Brain. *Stroke* 46:2293-2298.
- Korytkowski M, Thomas A, Reid L, Tedesco MB, Gooding WE, Gerich J (2002) Glimepiride improves both first and second phases of insulin secretion in type 2 diabetes. *Diabetes Care* 25:1607-1611.
- Kottenberg E, Thielmann M, Bergmann L, Heine T, Jakob H, Heusch G, Peters J (2012) Protection by remote ischemic preconditioning during coronary artery bypass graft surgery with isoflurane but not propofol - a clinical trial. *Acta Anaesthesiol Scand* 56:30-38.
- Kramer W, Muller G, Girbig F, Gutjahr U, Kowalewski S, Hartz D, Summ HD (1995) The molecular interaction of sulfonylureas with beta-cell ATP-sensitive K(+)-channels. *Diabetes Res Clin Pract* 28 Suppl:S67-80.
- Kristiansen SB, Lofgren B, Stottrup NB, Khatir D, Nielsen-Kudsk JE, Nielsen TT, Botker HE, Flyvbjerg A (2004) Ischaemic preconditioning does not protect the heart in obese and lean animal models of type 2 diabetes. *Diabetologia* 47:1716-1721.
- Langtry HD, Balfour JA (1998) Glimepiride. A review of its use in the management of type 2 diabetes mellitus. *Drugs* 55:563-584.
- Lee HT, Emala CW (2000) Protective effects of renal ischemic preconditioning and adenosine pretreatment: role of A(1) and A(3) receptors. *Am J Physiol Renal Physiol* 278:F380-387.
- Lennon SL, Quindry J, Hamilton KL, French J, Staib J, Mehta JL, Powers SK (2004) Loss of exercise-induced cardioprotection after cessation of exercise. *J Appl Physiol* (1985) 96:1299-1305.
- Leon BM, Maddox TM (2015) Diabetes and cardiovascular disease: Epidemiology, biological mechanisms, treatment recommendations and future research. *World J Diabetes* 6:1246-1258.
- Lloris-Carsi JM, Cejalvo D, Toledo-Pereyra LH, Calvo MA, Suzuki S (1993) Preconditioning: effect upon lesion modulation in warm liver ischemia. *Transplant Proc* 25:3303-3304.
- Luca MC, Liuni A, Muxel S, Munzel T, Forconi S, Gori T, Parker JD (2011) Chronic pharmacological preconditioning against ischemia. *Clin Hemorheol Microcirc* 49:287-293.
- Lucchinetti E, Bestmann L, Feng J, Freidank H, Clanachan AS, Finegan BA, Zaugg M (2012) Remote ischemic preconditioning applied during isoflurane inhalation provides no benefit to the myocardium of patients undergoing on-pump coronary artery bypass graft surgery: lack of synergy or evidence of antagonism in cardioprotection? *Anesthesiology* 116:296-310.
- Marso SP, Miller T, Rutherford BD, Gibbons RJ, Qureshi M, Kalynych A, Turco M, Schultheiss HP, Mehran R, Krucoff MW, Lansky AJ, Stone GW (2007) Comparison of myocardial reperfusion in patients undergoing percutaneous coronary intervention in ST-segment elevation acute myocardial infarction with versus without diabetes mellitus (from the EMERALD Trial). *Am J Cardiol* 100:206-210.
- Martin-Timon I, Sevillano-Collantes C, Segura-Galindo

- A, Del Canizo-Gomez FJ (2014) Type 2 diabetes and cardiovascular disease: Have all risk factors the same strength? *World J Diabetes* 5:444-470.
- Masada T, Xi G, Hua Y, Keep RF (2000) The effects of thrombin preconditioning on focal cerebral ischemia in rats. *Brain Res* 867:173-179.
- Matsumoto S, Cho S, Tosaka S, Ureshino H, Maekawa T, Hara T, Sumikawa K (2009) Pharmacological preconditioning in type 2 diabetic rat hearts: the roles of mitochondrial ATP-sensitive potassium channels and the phosphatidylinositol 3-kinase-Akt pathway. *Cardiovasc Drugs Ther* 23:263-270.
- McCrinkle BW, Clarizia NA, Khaikin S, Holtby HM, Manlihot C, Schwartz SM, Caldarone CA, Coles JG, Van Arsdell GS, Scherer SW, Redington AN (2014) Remote ischemic preconditioning in children undergoing cardiac surgery with cardiopulmonary bypass: a single-center double-blinded randomized trial. *J Am Heart Assoc* 3.
- McGinnis GR, Ballmann C, Peters B, Nanayakkara G, Roberts M, Amin R, Quindry JC (2015) Interleukin-6 mediates exercise preconditioning against myocardial ischemia reperfusion injury. *Am J Physiol Heart Circ Physiol* 308:H1423-1433.
- Miki T, Nagashima K, Tashiro F, Kotake K, Yoshitomi H, Tamamoto A, Gono T, Iwanaga T, Miyazaki J, Seino S (1998) Defective insulin secretion and enhanced insulin action in KATP channel-deficient mice. *Proc Natl Acad Sci U S A* 95:10402-10406.
- Miura T, Goto M, Miki T, Sakamoto J, Shimamoto K, Imura O (1995) Glibenclamide, a blocker of ATP-sensitive potassium channels, abolishes infarct size limitation by preconditioning in rabbits anesthetized with xylazine/pentobarbital but not with pentobarbital alone. *J Cardiovasc Pharmacol* 25:531-538.
- Moreno PR, Murcia AM, Palacios IF, Leon MN, Bernardi VH, Fuster V, Fallon JT (2000) Coronary composition and macrophage infiltration in atherectomy specimens from patients with diabetes mellitus. *Circulation* 102:2180-2184.
- Moretti C et al. (2018) The EUROpean and Chinese cardiac and renal Remote Ischemic Preconditioning Study (EURO-CRIPS CardioGroup I): A randomized controlled trial. *Int J Cardiol* 257:1-6.
- Mounsey RA, Pang CY, Forrest C (1992) Preconditioning: a new technique for improved muscle flap survival. *Otolaryngol Head Neck Surg* 107:549-552.
- Murry CE, Jennings RB, Reimer KA (1986) Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. *Circulation* 74:1124-1136.
- National Eye Institute (2015) Facts About Diabetic Eye Disease. In, p <http://www.nei.nih.gov/health/diabetic/retinopathy>.
- Otsuka F NM, Sakakura K, Ladich E, Kolodgie FD, Virmani R, (2012) Pathology of Diabetic Atherosclerosis: Composition, Characteristics, and Distribution. In: *Diabetes in Cardiovascular Disease: A Companion to Braunwald's Heart Disease, 9th Edition* (McGuire DK MN, ed), pp 87-98: ELSEVIER Saunders.
- Ottenbacher KJ, Ostir GV, Peek MK, Markides KS (2004) Diabetes mellitus as a risk factor for stroke incidence and mortality in Mexican American older adults. *J Gerontol A Biol Sci Med Sci* 59:M640-645.
- Ovunc K (2000) Effects of glibenclamide, a K(ATP) channel blocker, on warm-up phenomenon in type II diabetic patients with chronic stable angina pectoris. *Clin Cardiol* 23:535-539.
- Ozbilgin S, Ozkardesler S, Akan M, Boztas N, Ozbilgin M, Ergur BU, Derici S, Guneli ME, Meseri R (2016) Renal Ischemia/Reperfusion Injury in Diabetic Rats: The Role of Local Ischemic Preconditioning. *Biomed Res Int* 2016:8580475.
- Pajdo R, Brzozowski T, Konturek PC, Kwiecien S, Konturek SJ, Sliwowski Z, Pawlik M, Ptak A, Drozdowicz D, Hahn EG (2001) Ischemic preconditioning, the most effective gastroprotective intervention: involvement of prostaglandins, nitric oxide, adenosine and sensory nerves. *Eur J Pharmacol* 427:263-276.
- Pickard JM et al. (2015) Remote ischemic conditioning: from experimental observation to clinical application: report from the 8th Biennial Hatter Cardiovascular Institute Workshop. *Basic Res Cardiol* 110:453.
- Plamondon H, Blondeau N, Heurteaux C, Lazdunski M (1999) Mutually protective actions of kainic acid epileptic preconditioning and sublethal global ischemia on hippocampal neuronal death: involvement of adenosine A1 receptors and K(ATP) channels. *J Cereb Blood Flow Metab* 19:1296-1308.
- Plutzky J ZB, Brown JD, (2012) Vascular Biology of Atherosclerosis in Patients with Diabetes: Dyslipidemia, Hypercoagulability, Endothelial Dysfunction and Inflammation. In: *Diabetes in Cardiovascular Disease: A Companion to Braunwald's Heart Disease, 9th Edition* (McGuire DK MN, ed), pp 111-126: ELSEVIER Saunders.
- Pop-Busui R, Boulton AJ, Feldman EL, Bril V, Freeman R, Malik RA, Sosenko JM, Ziegler D (2017) Diabetic Neuropathy: A Position Statement by the American Diabetes Association. *Diabetes Care* 40:136-154.
- Przyklenk K (2013) Reduction of myocardial infarct size with ischemic "conditioning": physiologic and technical considerations. *Anesth Analg* 117:891-901.
- Przyklenk K, Maynard M, Greiner DL, Whittaker P (2011) Cardioprotection with postconditioning: loss of efficacy in murine models of type-2 and type-1 diabetes. *Antioxid Redox Signal* 14:781-790.
- Przyklenk K, Bauer B, Ovize M, Kloner RA, Whittaker P (1993) Regional ischemic 'preconditioning' protects remote virgin myocardium from subsequent sustained coronary occlusion. *Circulation* 87:893-899.
- Quindry J, French J, Hamilton K, Lee Y, Mehta JL, Powers S (2005) Exercise training provides cardioprotection against ischemia-reperfusion induced apoptosis in young and old animals. *Exp Gerontol* 40:416-425.
- Rahmi RM, Uchida AH, Rezende PC, Lima EG, Garzillo CL, Favarato D, Strunz CM, Takiuti M, Girardi P, Hueb W, Kalil Filho R, Ramires JA (2013) Effect of hypoglycemic agents on ischemic preconditioning in patients with type 2 diabetes and symptomatic coronary artery disease. *Diabetes Care* 36:1654-1659.
- Raphael J, Gozal Y, Navot N, Zuo Z (2010) Hyperglycemia inhibits anesthetic-induced postconditioning in the rabbit heart via modulation of phosphatidylinositol-3-kinase/Akt and endothelial nitric oxide synthase signaling. *J Cardiovasc Pharmacol* 55:348-357.
- Raval AP, Bramlett H, Perez-Pinzon MA (2006a) Estrogen preconditioning protects the hippocampal CA1 against ischemia. *Neuroscience* 141:1721-1730.
- Raval AP, Dave KR, Perez-Pinzon MA (2006b) Resveratrol mimics ischemic preconditioning in the brain. *J Cereb Blood Flow Metab* 26:1141-1147.
- Schramm TK, Gislason GH, Kober L, Rasmussen S, Rasmussen JN, Abildstrom SZ, Hansen ML, Folke F, Buch P, Madsen M, Vaag A, Torp-Pedersen C (2008) Diabetes patients requiring glucose-lowering therapy and nondiabetics with a prior myocardial infarction carry the same cardiovascular risk: a population study of 3.3 million people. *Circulation* 117:1945-1954.

- Schulz R, Rose J, Heusch G (1994) Involvement of activation of ATP-dependent potassium channels in ischemic preconditioning in swine. *Am J Physiol* 267:H1341-1352.
- Sengul I, Sengul D, Guler O, Hasanoglu A, Urhan MK, Taner AS, Vinten-Johansen J (2013) Postconditioning attenuates acute intestinal ischemia-reperfusion injury. *Kaohsiung J Med Sci* 29:119-127.
- Shi H, Sun BL, Zhang J, Lu S, Zhang P, Wang H, Yu Q, Stetler RA, Vosler PS, Chen J, Gao Y (2013) miR-15b suppression of Bcl-2 contributes to cerebral ischemic injury and is reversed by sevoflurane preconditioning. *CNS Neurol Disord Drug Targets* 12:381-391.
- Sivaraman V, Pickard JM, Hausenloy DJ (2015) Remote ischaemic conditioning: cardiac protection from afar. *Anaesthesia* 70:732-748.
- Spencer EA, Pirie KL, Stevens RJ, Beral V, Brown A, Liu B, Green J, Reeves GK, Million Women Study Collaborators (2008) Diabetes and modifiable risk factors for cardiovascular disease: the prospective Million Women Study. *Eur J Epidemiol* 23:793-799.
- Staat P, Rioufol G, Piot C, Cottin Y, Cung TT, L'Huillier I, Aupetit JF, Bonnefoy E, Finet G, Andre-Fouet X, Ovize M (2005) Postconditioning the human heart. *Circulation* 112:2143-2148.
- Stagliano NE, Perez-Pinzon MA, Moskowitz MA, Huang PL (1999) Focal ischemic preconditioning induces rapid tolerance to middle cerebral artery occlusion in mice. *J Cereb Blood Flow Metab* 19:757-761.
- Sun K, Liu ZS, Sun Q (2004) Role of mitochondria in cell apoptosis during hepatic ischemia-reperfusion injury and protective effect of ischemic postconditioning. *World J Gastroenterol* 10:1934-1938.
- Tai W, Shi E, Yan L, Jiang X, Ma H, Ai C (2012) Diabetes abolishes the cardioprotection induced by sevoflurane postconditioning in the rat heart in vivo: roles of glycogen synthase kinase-3beta and its upstream pathways. *J Surg Res* 178:96-104.
- Tanaka K, Kehl F, Gu W, Krolikowski JG, Pagel PS, Warltier DC, Kersten JR (2002) Isoflurane-induced preconditioning is attenuated by diabetes. *Am J Physiol Heart Circ Physiol* 282:H2018-2023.
- Theodoraki K, Karmanioliou I, Tympa A, Tasoulis MK, Nastos C, Vassiliou I, Arkadopoulou N, Smyrniotis V (2016) Beyond Preconditioning: Postconditioning as an Alternative Technique in the Prevention of Liver Ischemia-Reperfusion Injury. *Oxid Med Cell Longev* 2016:8235921.
- Thomaz Neto FJ, Koike MK, Abrahao Mde S, Carillo Neto F, Pereira RK, Machado JL, Montero EF (2013) Ischemic preconditioning attenuates remote pulmonary inflammatory infiltration of diabetic rats with an intestinal and hepatic ischemia-reperfusion injury. *Acta Cir Bras* 28:174-178.
- Tomai F, Crea F, Gaspardone A, Versaci F, De Paulis R, Penta de Peppo A, Chiariello L, Gioffre PA (1994) Ischemic preconditioning during coronary angioplasty is prevented by glibenclamide, a selective ATP-sensitive K<sup>+</sup> channel blocker. *Circulation* 90:700-705.
- Tosaki A, Engelman DT, Engelman RM, Das DK (1996) The evolution of diabetic response to ischemia/reperfusion and preconditioning in isolated working rat hearts. *Cardiovasc Res* 31:526-536.
- Tracey ML, Gilmartin M, O'Neill K, Fitzgerald AP, McHugh SM, Buckley CM, Canavan RJ, Kearney PM (2016) Epidemiology of diabetes and complications among adults in the Republic of Ireland 1998-2015: a systematic review and meta-analysis. *BMC Public Health* 16:132.
- Tsang A, Hausenloy DJ, Mocanu MM, Carr RD, Yellon DM (2005) Preconditioning the diabetic heart: the importance of Akt phosphorylation. *Diabetes* 54:2360-2364.
- Ueda K, Komine J, Matsuo M, Seino S, Amachi T (1999) Cooperative binding of ATP and MgADP in the sulfonylurea receptor is modulated by glibenclamide. *Proc Natl Acad Sci U S A* 96:1268-1272.
- Vinten-Johansen J, Shi W (2011) Preconditioning and postconditioning: current knowledge, knowledge gaps, barriers to adoption, and future directions. *J Cardiovasc Pharmacol Ther* 16:260-266.
- Wada K, Ito M, Miyazawa T, Katoh H, Nawashiro H, Shima K, Chigasaki H (1996) Repeated hyperbaric oxygen induces ischemic tolerance in gerbil hippocampus. *Brain Res* 740:15-20.
- Wang H, Lu S, Yu Q, Liang W, Gao H, Li P, Gan Y, Chen J, Gao Y (2011) Sevoflurane preconditioning confers neuroprotection via anti-inflammatory effects. *Front Biosci (Elite Ed)* 3:604-615.
- Xiong L, Lu Z, Hou L, Zheng H, Zhu Z, Wang Q, Chen S (2003) Pretreatment with repeated electroacupuncture attenuates transient focal cerebral ischemic injury in rats. *Chin Med J (Engl)* 116:108-111.
- Yadav HN, Singh M, Sharma PL (2010) Involvement of GSK-3beta in attenuation of the cardioprotective effect of ischemic preconditioning in diabetic rat heart. *Mol Cell Biochem* 343:75-81.
- Yue TL, Bao W, Gu JL, Cui J, Tao L, Ma XL, Ohlstein EH, Jucker BM (2005) Rosiglitazone treatment in Zucker diabetic fatty rats is associated with ameliorated cardiac insulin resistance and protection from ischemia/reperfusion-induced myocardial injury. *Diabetes* 54:554-562.
- Zhao H (2007) The protective effect of ischemic postconditioning against ischemic injury: from the heart to the brain. *J Neuroimmune Pharmacol* 2:313-318.
- Zhao H (2009) Ischemic postconditioning as a novel avenue to protect against brain injury after stroke. *J Cereb Blood Flow Metab* 29:873-885.
- Zhao H, Sapolsky RM, Steinberg GK (2006) Interrupting reperfusion as a stroke therapy: ischemic postconditioning reduces infarct size after focal ischemia in rats. *J Cereb Blood Flow Metab* 26:1114-1121.
- Zhao H, Ren C, Chen X, Shen J (2012) From rapid to delayed and remote postconditioning: the evolving concept of ischemic postconditioning in brain ischemia. *Curr Drug Targets* 13:173-187.
- Zhao ZQ, Corvera JS, Halkos ME, Kerendi F, Wang NP, Guyton RA, Vinten-Johansen J (2003) Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 285:H579-588.
- Zhu SG, Xi L, Kukreja RC (2012) Type 2 diabetic obese db/db mice are refractory to myocardial ischaemic post-conditioning in vivo: potential role for Hsp20, F1-ATPase delta and Echs1. *J Cell Mol Med* 16:950-958.
- Zhu XC, Jiang T, Zhang QQ, Cao L, Tan MS, Wang HF, Ding ZZ, Tan L, Yu JT (2015) Chronic Metformin Preconditioning Provides Neuroprotection via Suppression of NF-kappaB-Mediated Inflammatory Pathway in Rats with Permanent Cerebral Ischemia. *Mol Neurobiol* 52:375-385.